

5-Lipoxygenase inhibition by *N*-hydroxycarbamates in dual-function compounds

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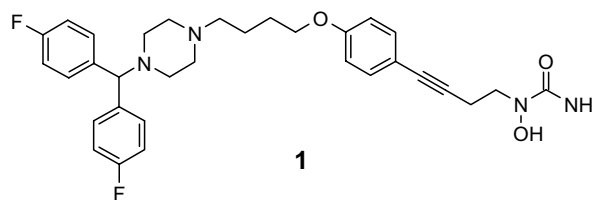
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Abstract—A series of *N*-hydroxycarbamates containing a histaminergic H₁ receptor antagonist pharmacophore was synthesized. In vitro assays determined the compounds had both histaminergic binding and 5-lipoxygenase inhibiting activities comparable to the corresponding *N*-hydroxyurea analog. Animal models demonstrated antihistaminergic and the 5-lipoxygenase inhibitory activity, with the *N*-hydroxyurea analog having a better overall profile.

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Histamine plays a role in the pathophysiology of asthma alongside its key role in allergic response. Clinical studies have demonstrated that asthmatics treated simultaneously with an H₁ receptor antagonist and a LTD₄ receptor antagonist experienced less airway obstruction than those patients treated with either drug alone.¹ A similar combination proved to be as efficacious as a corticosteroid in treating the airway symptoms of allergic rhinitis and asthma.² Inhibition of 5-lipoxygenase (5-LO) is potentially a more efficacious treatment than antagonism of leukotriene receptors, as biosynthesis of all the pro-inflammatory leukotrienes would be diminished.

Our work has focused on dual-function antihistaminergic/5-LO inhibiting compounds such as the *N*-hydroxyurea UCB 62045 (**1**), which demonstrates both activities in vitro and in vivo.³ UCB 62045 is effective in reducing ovalbumin-induced bronchoconstriction in the guinea pig bronchoconstriction model with oral dosing, and has been investigated as a treatment for asthma and allergic rhinitis. Reports on the 5-LO inhibitory effect of *N*-hydroxycarbamates⁴ supported investigating this moiety in similar dual-function molecules.

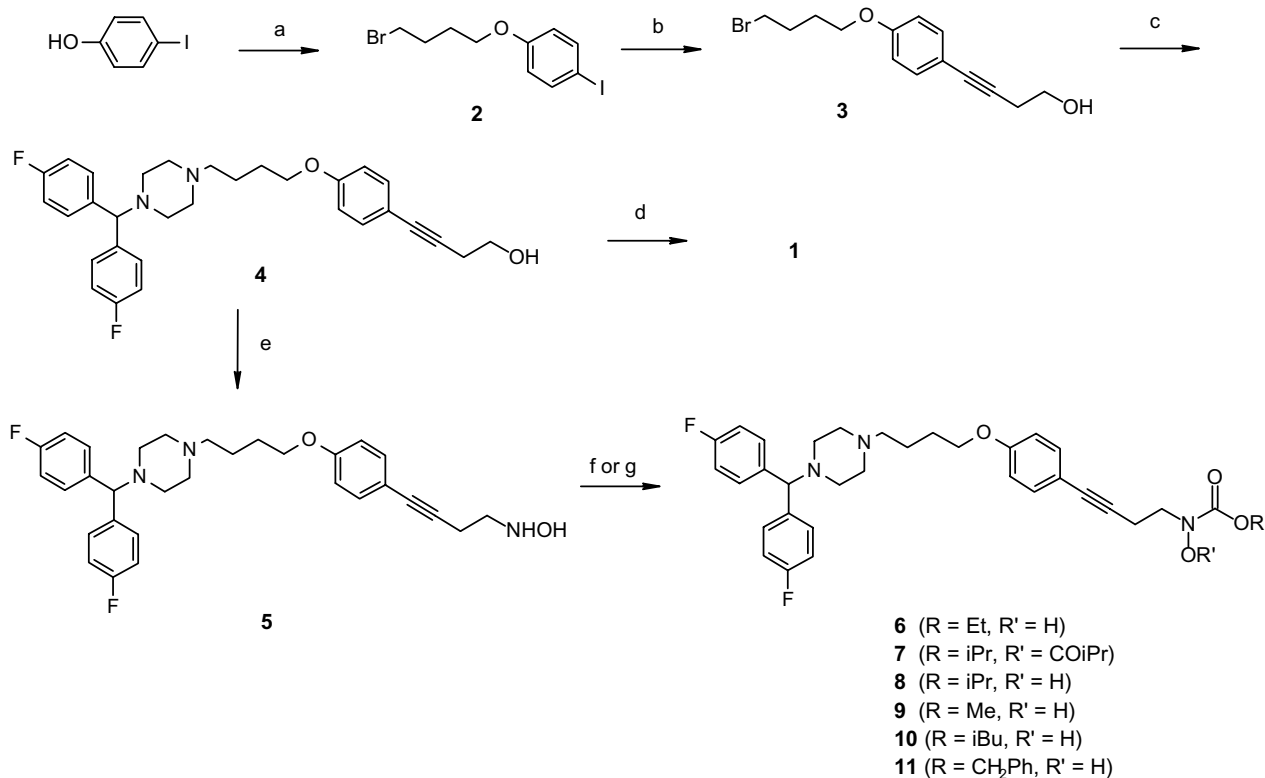


Synthesis of **1** began by reacting 4-iodophenol with 1,4-dibromobutane to give **2** (Scheme 1). Palladium catalyzed coupling of **2** with 3-butyn-1-ol gave **3**. *N*-Alkylation of 1-bis-(4-fluorophenyl)methyl piperazine with **3** gave alcohol **4**, which was converted to *N*-hydroxyurea **1** using a literature procedure.⁵ Synthesis of the carbamate analogs began with alcohol **4**, which was converted into hydroxylamine **5** via mesylation and displacement with aqueous hydroxylamine. Treatment of **5** with ethyl chloroformate gave the desired *N*-hydroxycarbamate **6**. However, treatment of **5** with isopropyl chloroformate gave predominately the undesired *N,O*-bis-acylated product **7** in 43% yield. Treatment of **7** with KOH/*i*-PrOH hydrolyzed only the carbonate group to afford *N*-hydroxycarbamate **8**. This two step procedure, alkyl chloroformate addition to **5** making the diacyl product followed by basic hydrolysis, also proved more convenient for the preparation of the methyl, isobutyl, and benzyl *N*-hydroxycarbamates **9–11**.⁶

Human H₁ receptor binding was performed using CHO-K1 cells expressing the recombinant H₁ receptor⁷ (Table

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Scheme 1. Reagents and conditions: (a) 1,4-dibromobutane, K₂CO₃/DMF (83%); (b) 3-butyne-1-ol, (Ph₃P)₂PdCl₂, CuI, Et₃N/CH₂Cl₂ (76%); (c) 1-[bis-(4-fluorophenyl)methyl]piperazine, K₂CO₃/DMF (77%); (d) (1) PPh₃, DIAD, PhOCO₂NHCOOPh/THF; (2) NH₃/MeOH (25%); (e) (1) MeSO₂Cl, Et₃N/CH₂Cl₂; (2) NH₂OH (aq), THF/MeOH (39%); (f) for **6**: EtOCOCl, Et₃N/CH₂Cl₂ (39%); (g) (1) ROCOCl, Et₃N/CH₂Cl₂; (2) KOH/ROH (**8**—21%, **9**—23%, **10**—32%, **11**—15%, two steps).

1). Hydroxyurea **1** displayed the best binding,⁸ better than the reference standard, cetirizine. In the carbamate series, the compounds with the smaller alkoxy groups (OR) displayed better H₁ binding, the methyl carbamate **9** binding better than cetirizine, the ethyl carbamate **6** binding equally, and all others binding with lower affinity. The overall variation in H₁ binding was not pronounced throughout the entire series.

Inhibition of 5-LO was tested with human recombinant 5-LO (HR),⁹ and a human whole blood (HWB) assay, which monitors calcium ionophore-induced LTB₄ for-

mation¹⁰ (Table 1). All the compounds tested were more potent than zileuton, the reference standard, with little variation occurring between the carbamate analogs.

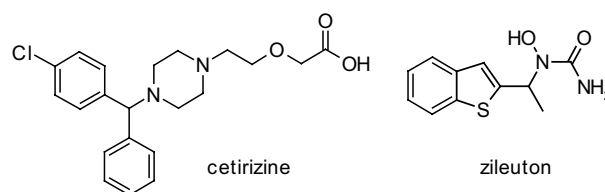


Table 1. In vitro activities of dual-function compounds^a

Compound	H ₁ binding (K _i nM) ^b	5-LO inhibition (HR, IC ₅₀ , nM) ^c	5-LO inhibition (HWB, IC ₅₀ , nM) ^d
1	3.63 ± 0.52	193 ± 76	89 ± 34
6	5.89 ± 0.49	365 ± 55	72 ± 40
8	9.55 ± 0.62	335 ± 115	102 ± 69
9	5.01 ± 0.59	170 ± 60	90 ± 11
10	7.59 ± 0.66	420 ± 30	176 ± 116
11	17.8 ± 0.60	350 ± 20	150 ± 56
Cetirizine	5.89 ± 0.81	—	—
Zileuton	—	518 ± 146 ^b	873 ± 391 ^{b,c}

^a Mean ± SD.

^b n ≥ 6.

^c n ≥ 2.

^d n ≥ 3.

^e Lit. value: 900.¹⁰

N-Hydroxyurea **1** and N-hydroxycarbamates **6** and **8** were tested in animal models with oral dosing (2 mg/kg). While methyl carbamate **9** was more potent in the H₁ and 5-LO (HR) assays, analogs **6** and **8** were expected to have better metabolic stability. Compounds **6** and **8** are both orally active antihistamines with 5-LO inhibitory activity for up to 6 h.

The H₁ antagonist activity was determined using the Konzett–Rössler protocol, which monitors histamine-induced bronchoconstriction (Fig. 1).¹¹ Antihistaminergic activity profiles of the three compounds were different; hydroxyurea **1**, after a slow onset, displayed nearly complete inhibition of bronchoconstriction at 3 and 6 h. Ethyl carbamate **6** had a faster onset and increasing activity with time, while isopropyl carbamate **8** had slow onset and the least activity at 3 and 6 h. None of the

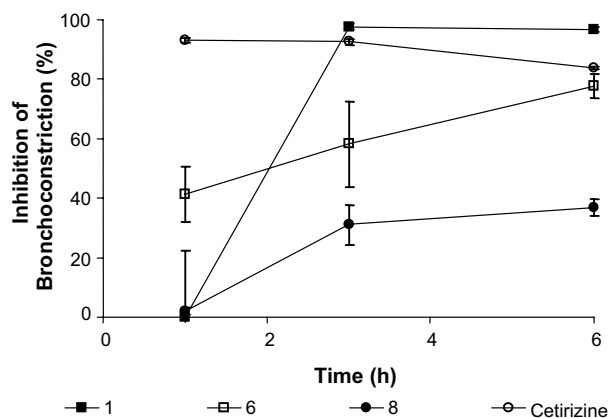


Figure 1. The effect of dual-function compounds on histamine-induced bronchoconstriction in guinea pigs (**1**, **6**, and **8**, 2 mg/kg, po; cetirizine, 0.5 mg/kg, po, $n = 3$ animals/timepoint).

dual-function compounds are as active as cetirizine, which displayed nearly complete inhibition of histamine-induced bronchoconstriction at all time points with a lower dose (0.5 mg/kg).

The ex vivo 5-LO assay monitors the inhibition of LTB₄ production after calcium ionophore-induced stimulation¹² (Fig. 2). Zileuton was tested alongside the dual-function compounds as a reference. Inhibition of 5-LO activity by the three dual-function molecules was detected up to six hours after challenge. *N*-Hydroxyurea **1** has better activity than the two *N*-hydroxycarbamates tested, and the activity increases with time. The *N*-hydroxycarbamate activities are fairly constant in this assay, or decrease with time.

In conclusion, in a dual-function antihistaminergic system, *N*-hydroxycarbamates demonstrate 5-LO inhibitory activity, both in vitro and in vivo with oral dosing. However, the ex vivo 5-LO activities of the *N*-hydroxycarbamates are lower at 1, 3, and 6 h than the analogous *N*-hydroxyurea. Further work is required to

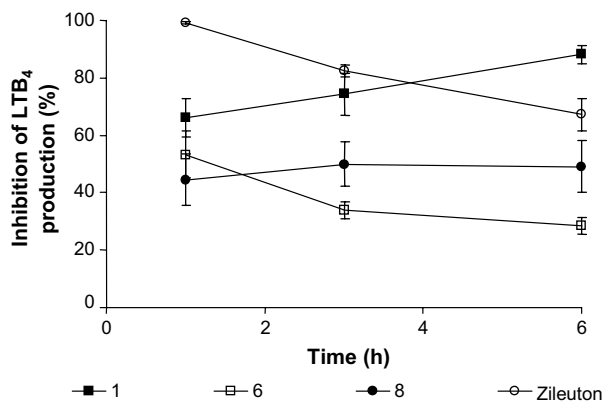


Figure 2. The effect of dual-function compounds and of zileuton on the inhibition of A-23187-stimulated LTB₄ production in guinea pigs (for **1**, **6**, **8**, 2 mg/kg, po; zileuton, 5 mg/kg, po, $n = 3$ animal/timepoint).

fully determine the therapeutic potential of *N*-hydroxycarbamates.¹³

Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2004.12.023.

References and notes

- Roquet, A.; Dahlen, B.; Kumlin, M.; Ihre, E.; Gudrun, A.; Binks, S. *Am. J. Resp. Crit. Care Med.* **1997**, *155*, 1856.
- Wilson, A. M.; Orr, L. C.; Sims, E. J.; Dempsey, O. J.; Lipworth, B. J. *Am. J. Resp. Crit. Care Med.* **2000**, *162*, 1297.
- Lewis, T. A.; Bayless, L.; Eckman, J. B.; Ellis, J. L.; Grewal, G.; Libertine, L.; Nicolas, J. M.; Scannell, R. T.; Wels, B. F.; Wenberg, K.; Wypij, D. M. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 2265; Lewis, T. A.; Young, M. A.; Arrington, M. P.; Bayless, L.; Cai, X.; Collart, P.; Eckman, J. B.; Ellis, J. L.; Ene, D. G.; Libertine, L.; Nicolas, J.-M.; Scannell, R. T.; Wels, B. F.; Wenberg, K.; Wypij, D. M. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 5591; Selig, W. M.; Bayless, L.; Libertine, L.; Eckman, J. B.; Wypij, D. M.; Wels, B. F.; Eckert, M.; Young, M. A.; Nicolas, J.-M.; Scannell, R. T.; Ellis, J. L. *Chest* **2003**, *123*, 371; Scannell, R. T. et al. *Inflamm. Res.* **2004**, *53*(Supplement 1), S33.
- Surman, M. D.; Mulvihill, M. J.; Miller, M. J. *J. Org. Chem.* **2002**, *67*, 4115; Yatabe, T.; Kawai, Y.; Oku, T.; Tanaka, H. *Chem. Pharm. Bull.* **1998**, *46*, 966; Connolly, P. J.; Wetter, S. K.; Beers, K. N.; Hamel, S. C.; Chen, R. H. K.; Wachter, M. P.; Ansell, J.; Singer, M. M.; Steber, M.; Ritchie, D. M.; Argentieri, D. C. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 979.
- Stewart, A. O.; Brooks, D. W. *J. Org. Chem.* **1992**, *57*, 5020.
- All compounds tested were characterized by ¹H NMR, mass spectral, and IR analysis; purity was >95% as determined by HPLC analysis with UV detection at 254 nm and tandem mass spectral detection. Recrystallization from EtOAc gave analytically pure material as determined by combustion analysis (C, H, N ± 0.4% of calculated values).
- Gillard, M.; Van Der Perren, C.; Moguelevsky, N.; Massingham, R.; Chatelain, P. *Mol. Pharmacol.* **2002**, *61*, 391.
- The discrepancy between the H₁ value for **1** reported here and in Refs. **3b,c** is due to testing at two different laboratories using different cell lines with different batches of **1**.
- Brooks, C. D.; Stewart, A. O.; Basha, A.; Bhatia, P.; Ratajczyk, J. D.; Martin, J. G.; Craig, R. A.; Kolasa, T.; Bouska, J. B.; Lanni, C.; Harris, R. R.; Malo, P. E.; Carter, G. C.; Bell, R. L. *J. Med. Chem.* **1995**, *38*, 4768.
- Carter, G. W.; Young, P. R.; Albert, D. H.; Bouska, J.; Dyer, R.; Bell, R. L.; Summers, J. B.; Brooks, D. W. *J. Pharmacol. Exp. Ther.* **1991**, *256*, 929.
- Konzett, H.; Rössler, R. *Naodyn-Schmiedebergs Arch. Exp. Pathol. Pharmacol.* **1940**, *195*, 71.
- Spaethe, S. M.; Snyder, D. W.; Pechous, P. A.; Clarke, T.; VanAlstyne, E. L. *Biochem. Pharmacol.* **1992**, *43*, 377.
- Experimental procedures for the synthesis of all intermediates and final compounds, plus the biological experimental procedures can be found in the [Supplementary material](#).